

Netamines O–S, five new tricyclic guanidine alkaloids from the Madagascar sponge Biemna laboutei and their antimalarial activity.

Emmanuelle Gros, Marie-Therese Martin, Jonathan Sorres, Céline Moriou, Jean Vacelet, Michel Frederich, Maurice Aknin, Yoel Kashman, Anne Gauvin-Bialecki, Ali Al Mourabit

▶ To cite this version:

Emmanuelle Gros, Marie-Therese Martin, Jonathan Sorres, Céline Moriou, Jean Vacelet, et al.. Netamines O–S, five new tricyclic guanidine alkaloids from the Madagascar sponge Biemna laboutei and their antimalarial activity.. Chemistry and Biodiversity, 2015, 12, pp.1725-1733. 10.1002/cbdv.201400350. hal-01243705

HAL Id: hal-01243705 https://hal.univ-reunion.fr/hal-01243705

Submitted on 16 Nov 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Netamines O-S, Five New Tricyclic Guanidine Alkaloids from the Madagascar Sponge *Biemna laboutei*, and Their Antimalarial Activities

by Emmanuelle Gros^a), Marie-Thérèse Martin^b), Jonathan Sorres^b), Céline Moriou^b), Jean Vacelet^c), Michel Frederich^d), Maurice Aknin^a), Yoel Kashman^c), Anne Gauvin-Bialecki^{*a}), and Ali Al-Mourabit^b)

^a) Laboratoire de Chimie des Substances Naturelles et des Sciences des Aliments, Faculté des Sciences et Technologies, Université de La Réunion, 15 Avenue René Cassin, CS 92003, FR-97744 Saint-Denis Cedex 9, La Réunion

(phone: +262-262-938197; fax: +262-262-938183; e-mail: anne.bialecki@univ-reunion.fr) ^b) Centre de Recherche de Gif-sur-Yvette, Institut de Chimie des Substances Naturelles, UPR 2301, CNRS, Avenue de la Terrasse, FR-91198 Gif-sur-Yvette

, Avenue de la Terrasse, FR-91198 Oil-sui-

^c) Aix Marseille Université, CNRS, IRD, Avignon Université, IMBE UMR 7263, Station marine d'Endoume, FR-13397 Marseille

^d) Laboratory of Pharmacognosy, Department of Pharmacy, CIRM, University of Liège B36, BE-4000 Liège

e) School of Chemistry, Sackler Faculty of Exact Sciences, Tel Aviv University, Tel Aviv 69978, Israel

In our continuing program to isolate new compounds from the Madagascar sponge *Biemna laboutei*, five new tricyclic guanidine alkaloids, netamines O – S (1–5, resp.), have been identified together with the known compounds netamine E (6) and mirabilin J (7). The structures of all new netamines were assigned on the basis of spectroscopic analyses. Their relative configurations were established by analysis of ROESY data and comparison with literature data. Netamines O, P, and Q, which were isolated in sufficient quantities, were tested for their cytotoxic activities against KB cells and their activities against the malaria parasite *Plasmodium falciparum*. Netamines O and Q were found to be moderately cytotoxic. Netamines O, P, and Q exhibited antiplasmodial activities with IC_{50} values of 16.99 ± 4.12 , 32.62 ± 3.44 , and $8.37 \pm 1.35 \mu$ M, respectively.

Introduction. – Tricyclic guanidine compounds, bearing a tricyclic 2-amino-1,3diazaoctahydroacenaphtylene skeleton, like ptilocaulins, mirabilins, and netamines, are restricted to marine sponges of the order Poecilosclerida [1–11] and can be grouped on the basis of their degree of oxidation and C=C bond position with pyrimidine, $\Delta^{7.8-}$, $\Delta^{8.8a-}$, $\Delta^{8a,8b}$ -unsaturated, or saturated heterocycles. Many of these alkaloids were reported to possess noteworthy biological activities, *i.e.*, cytotoxic [1][4][8][9][12], antibacterial [1][7], antifungal [3], antimalarial [3][11], and antiprotozoal activities [3].

In 2006, we reported the isolation and structure elucidation of seven alkaloids, named netamines A–G, from the Madagascar sponge *Biemna laboutei* HOOPER, 1996 collected twice in May 2004 near Sainte Marie Island, and once in January 2005 at Itampule [9]. From yet another collection of the sponge in October 2009 in Salary Bay (*ca.* 100 km north of Tulear), we isolated several additional tricyclic guanidine alkaloids, including the known netamine G, mirabilins A, C, and F, and seven new

tricyclic alkaloids, netamines H-N [11]. All the latter new compounds isolated from *B. laboutei* possess a 5,6,8b-triazaperhydroacenaphtylene skeleton. Except for netamines A-D that possess a saturated ring system, netamine E shows a C(8a)=C(8b) bond, netamines K-N have a C(8)=C(8a) bond, and netamines F-J contain a pyrimidine ring system.

In this article, we describe the isolation, structure elucidation, and biological characterization of a series of five new tricyclic guanidines, netamines O-S (1-5), along with the known compounds netamine E and mirabilin J. All these compounds, isolated from the 2009 Salary Bay sponge, show a C(8a)=C(8b) bond.

Results and Discussion. – *Structure Elucidation.* Chemical fractionation of the crude extract, using a sequence of medium pressure liquid chromatography over silica gel or reversed phase, followed by repetitive reversed-phase preparative HPLC, yielded new (1–5) and known (6 and 7) compounds. The known compounds, netamine E (6) and mirabilin J (7) were identified by comparison with published spectroscopic data [8][9] revised by *Snider* [10]. For the known isolated compound netamine E (6), the $[\alpha]_D^{25}$ value of +37.0 ($[\alpha]_D^{25} = +35.0$ [9]) establishes the absolute configuration as shown in the *Figure*. Although the optical rotations of all co-isolated compounds have the same algebraic sign as netamine E, we were careful not to conclude for compounds 1-5 and favored to stick to our ROESY data and the relative configuration.

Netamine O (1) was obtained as pale yellow oil. The molecular formula was established by HR-ESI-MS to be $C_{17}H_{28}N_3$. Analysis of the 1D and 2D ¹H-, ¹³C-, and ¹⁵N-NMR data for 1 (*Table 1*) revealed resonances and correlations consistent with those of a tricyclic guanidinium system with five C=C bond equivalents, three CH₂ (δ (C) 30.9, 34.1, and 34.2) and four CH groups (34.8, 38.8, 39.9, and 53.0), a tetra-

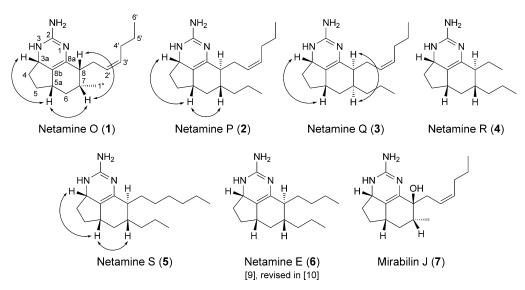


Figure. Structures of the tricyclic guanidine alkaloids netamines O-S (1–5, resp.), netamine E (6), and mirabilin J (7). Relative configurations were determined by ¹H,¹H-ROESY (H \leftrightarrow H) correlations.

Position	$\delta(\mathrm{H})$	$\delta(C)$	COSY	HMBC $(H \rightarrow C)$
1	12.10 ^a)	_	_	_
2	_	154.3	-	_
$H_2N-C(2)$	6.49 ^a)	_	_	-
3	9.85 ^a)	_	-	-
3a	4.23 - 4.28 (m)	53.0	-	5, 8b
4	2.23-2.25(m)	34.1	5	8b
5	1.18 - 1.21 (m), 1.70 - 1.76 (m)	34.2	4, 5a	3a, 8b
5a	2.46-2.52(m)	38.8	5, 6	5, 8a, 8b
6	1.21 - 1.23 (m)	30.9	5a, 7	-
7	1.98-2.05(m)	34.8	1", 6	1', 1"
8	2.31-2.35(m)	39.9	-	1', 1"
8a	_	129.4	_	-
8b	_	120.6	_	-
1′	2.26-2.30(m)	26.9	8	2′, 8, 8a
2′	5.34-5.41(m)	129.1	3′	3'
3′	5.37 - 5.43 (m)	131.8	2', 4'	4', 5'
4′	1.96-2.01(m)	30.7	3', 5'	3', 6'
5'	1.34 - 1.41 (m)	23.9	4', 6'	3', 6'
6'	0.92(t, J=7.1)	14.3	5'	4', 5'
1″	1.07 (d, J = 7.0)	19.4	7	7,8

Table 1. ¹*H*- and ¹³*C*-*NMR Data* (500 and 125 MHz, resp.; in CD₃OD) for Netamine O (1). δ in ppm, *J* in Hz.

substituted C=C bond (120.6 and 129.4), a guanidine-like C-atom (154.3), and three Natoms ($\delta(N)$ 78.1, 89.1, and 103.4). The HMB correlations of H–C(5a) and CH₂(1') with C(8a) (δ (C) 129.4) and of H–C(3a), CH₂(4), CH₂(5), and H–C(5a) with C(8b) (120.6) indicated a $\Delta^{8a,8b}$ -unsaturated tricyclic ring system like in netamine E [9]. In addition, HMBC data clearly indicated that the heterocyclic ring system is substituted at C(7) (δ (C) 34.8) and C(8) (39.9) by two alkyl groups summing up a total of seven Catoms. One substituent is a (2Z)-hex-2-en-1-yl group identified by a C(1') to C(6')correlation sequence. Its linkage to the C(8)-atom was determined by the HMB correlation of $CH_2(1')$ with C(8). The geometry of the C=C bond was confirmed by a NOE correlation between $CH_2(1')$ ($\delta(H)$ 2.26–2.30) and $CH_2(4')$ (1.96–2.01). Therefore, the second substituent has to be a Me group ($\delta(H)$ 1.07) attached to C(7) on the basis of the COSY correlation of Me(1'') with H-C(7). Moreover, the relative configuration of the four stereogenic centers at C(3a), C(5a), C(7), and C(8) was determined by ROESY correlations between H–C(3a) (δ (H) 4.23–4.28) and H–C(5a) (2.46–2.52), H–C(5a) and H–C(7) (1.98–2.05), and H–C(7) and H–C(8) (2.31–2.35). Thus, the four H-atoms were deduced to be at the same side of the tricyclic ring system and the two side chains were established to be cis to each other. Therefore, the relative configuration of Netamine O(1) is as shown in the *Figure*.

Netamine P (2) was isolated as white amorphous solid and has a molecular formula of $C_{19}H_{32}N_3$, which was suggested by HR-ESI-MS. Analysis of the NMR data (*Tables 2* and *3*) indicated again a guanidinium moiety, *i.e.*, the presence of three CH₂ (δ (C) 30.3,

Position	1 ^a)	2 ^a)	3 ^b)	4 ^a)	5 ^a)
3a	4.23-4.28 (<i>m</i>)	4.26-4.30 (<i>m</i>)	4.23-4.26 (<i>m</i>)	4.25-4.28 (<i>m</i>)	4.25-4.30 (m)
4	2.23 - 2.25(m)	1.65 - 1.69(m),	1.70 - 1.76(m),	1.65 - 1.72(m),	1.63 - 1.69(m)
		2.16 - 2.23(m)	2.22 - 2.25(m)	2.16 - 2.24(m)	2.16-2.23(m)
5	1.18 - 1.21 (m),	1.28 - 1.34(m),	1.20 - 1.23 (m)	1.22 - 1.29(m),	1.97 - 2.03(m)
	1.70 - 1.76(m)	1.96 - 2.02 (m)		1.98 - 2.03 (m)	
5a	2.46 - 2.52(m)	2.36 - 2.42(m)	2.44 - 2.50 (m)	2.32 - 2.38(m)	2.37 - 2.45(m)
6	1.21 - 1.23 (m)	2.09-2.13(m),	1.17 - 1.19(m),	0.74 - 0.79(m),	0.73 - 0.80 (m)
		0.73 - 0.80 (m)	1.78 - 1.83 (m)	2.06 - 2.12 (m)	2.09 - 2.14(m)
7	1.98 - 2.05(m)	1.52 - 1.56(m)	1.83 - 1.88(m)	1.53 - 1.59(m)	1.54 - 1.60 (m)
8	2.31 - 2.35(m)	1.97 - 2.00 (m)	2.38 - 2.41 (m)	1.93 - 1.99(m)	1.92 - 1.97(m)
1′	2.26 - 2.30(m)	2.30-2.36(m),	2.24 - 2.27 (m)	1.71 - 1.78(m)	1.57 - 1.65(m)
		2.52 - 2.58(m)			1.70 - 1.73 (m)
2′	5.34 - 5.41 (m)	5.27 - 5.33(m)	5.36 - 5.41 (m)	0.81 (t, J = 7.6)	1.15 - 1.20 (m)
	. ,				1.25 - 1.27 (m)
3′	5.37 - 5.43(m)	5.48 - 5.54(m)	5.37 - 5.43 (m)	_	1.27 - 1.34(m)
4′	1.96 - 2.01 (m)	2.04 - 2.08(m)	1.98 - 2.02 (m)	_	1.27 - 1.34(m)
5'	$1.34 - 1.41 \ (m)$	1.36 - 1.44(m)	$1.34 - 1.41 \ (m)$	_	1.27 - 1.34(m)
6'	0.92(t, J=7.1)	0.93(t, J=7.1)	0.91 (t, J = 7.3)	_	0.90(t, J = 6.9)
1″	1.07 (d, J = 7.0)	1.16 - 1.23(m),	1.30 - 1.32(m),	1.14 - 1.22 (m)	1.17 - 1.22 (m)
		1.60 - 1.63(m)	1.42 - 1.47 (m)		
2''	_	1.24 - 1.32(m),	1.40 - 1.42 (m)	1.27 - 1.35(m)	1.27 - 1.34(m)
		1.48 - 1.52 (m)			1.46 - 1.53 (m)
3''	_	0.94(t, J=7.0)	0.95(t, J = 6.9)	0.94(t, J=7.1)	0.94(t, J=7.1)

Table 2. ¹*H*-*NMR Data* (in CD₃OD) of Netamines O-S (1–5, resp.). δ in ppm, J in Hz.

34.0, and 36.3) and four CH groups (37.6, 38.9, 42.2, and 54.2), a tetrasubstituted C=C bond (120.0 and 128.6), a deshielded sp²-hybridized C-atom (159.7), and three N-atoms (δ (N) 78.1, 89.1, and 102.2). Relevant correlations between the ¹H- and ¹³C-NMR spectra for netamines O and P (**1** and **2**; resp.) supported a tricyclic guanidine-like arrangement with a C(8a)=C(8b) bond. However, the differences observed between **1** and **2** in the ¹H- and ¹³C-NMR spectra indicated that the Me group at C(7) of **1** is replaced by a Pr group at C(7) in **2**. The latter group was established by COSY correlations observed between Me(3'') (δ (H) 0.94), CH₂(2'') (1.24–1.32 and 1.48–1.52), CH₂(1'') (1.16–1.23 and 1.60–1.63), and H–C(7) (1.52–1.56). NOESY correlations between H–C(5a) and H–C(3a) and H–C(7) determined all three to be positioned at the same side of the molecule. In addition, NOESY correlations recorded in (D₇)DMF suggested that H–C(3a) (δ (H) 4.26–4.30), H–C(5a) (2.36–2.42), and H–C(7) (1.52–1.56) are located at the same side, while H–C(8) (1.97–2.00) is located at the opposite side of the molecule. Thus, the side chains of **2** are *trans* to each other.

Netamine Q (3) was obtained as white amorphous solid. The MS spectrum suggested the same formula $C_{19}H_{32}N_3$ as for 2, implying an isomeric structure. Comparison of their 1D- and 2D-NMR data, reinforced by a detailed analysis of the 2D-NMR spectra, also revealed that the two molecules have the same constitution and

Position	1 ^a)	2 ^a)	3 ^b)	4 ^a)	5 ^a)
2	154.3	159.7	154.4	154.8	154.7
3a	53.0	54.2	53.0	54.0	54.1
4	34.1	34.0	34.1	34.1	34.0
5	34.2	30.3	31.1	30.4	30.4
5a	38.8	37.6	38.9	37.6	37.7
6	30.9	36.3	32.5	36.5	36.6
7	34.8	38.9	38.4	37.7	38.4
8	39.9	42.2	40.2	42.9	42.3
8a	129.4	128.6	129.5	128.4	128.8
8b	120.6	120.0	120.7	120.0	119.7
1′	26.9	27.2	27.1	21.3	29.1
2′	129.1	125.9	129.0	8.5	25.2
3′	131.8	133.7	131.8	-	31.1
4′	30.7	30.9	30.7	-	33.1
5'	23.9	24.0	24.0	-	23.8
6'	14.3	14.3	14.3	-	14.5
1″	19.4	37.4	36.6	37.4	37.4
2''	-	21.6	21.8	21.4	21.4
3″	_	14.9	14.7	14.8	14.8

Table 3. ¹³C-NMR Data (in CD₃OD) of Netamines O-S (1–5, resp.). δ in ppm.

therefore different relative configurations. A NOESY experiment on **3** recorded in $(D_7)DMF$ revealed that H–C(3a) ($\delta(H)$ 4.23–4.26) and H–C(5a) (2.44–2.50) are located at the same side, whereas H–C(7) (1.83–1.88) and H–C(8) (2.38–2.41) appeared to be located at the opposite side of the molecule. The two side chains were therefore established to be *cis*, as in netamine O (**1**), but at the opposite side of the molecule compared to **1**. Additionally, as in netamines O (**1**) and P (**2**), the geometry of the C(2')=C(3') bond was assigned to be *cis* on the basis of the NOESY correlation between CH₂(1') and CH₂(4').

Netamine R (4) was isolated as pale yellow oil and showed the molecular composition of $C_{15}H_{26}N_3$, consistent with an ammonium salt with five C=C bond equivalents. Analysis of the 1D and 2D ¹H-, ¹³C-, and ¹⁵N-NMR data for 4 revealed resonances and correlations consistent with those of a tricyclic guanidine with a C(8a)=C(8b) bond. The HMB correlations for 4 clearly indicated that the heterocyclic ring system is substituted at C(7) (δ (C) 37.7) and C(8) (42.9) by a Pr and an Et group, resp. The position of the Pr group at C(7) was confirmed by COSY correlations between CH₂(1") and H–C(7), and HMB correlations of Me(3") with C(2") and C(1"). Likewise, the location of the Et group at C(8) was confirmed by the COSY correlations Me(2')/CH₂(1') and CH₂(1')/H–C(8). The absence of NOE correlations prevented the determination of the configuration of 4. The latter was deduced from empirical comparison of the chemical shifts of H–C(3a), H–C(5a), H–C(7), and H–C(8) belonging to 4 with those of 1–3 and 6. *Tables 2* and 3, set up for the purpose, showed that chemical shifts of the four CH H-atoms of netamine R were close to those of netamine E (6) [9][10].

Thus, it was finally assumed that, as for **2** and **6**, H-C(3a), H-C(5a), and H-C(7) are at the same side and H-C(8) is located at the opposite side of the molecule in **4**. The two side chains are therefore *trans*-oriented.

Netamine S (5) was isolated as brown oil. The molecular formula $C_{19}H_{34}N_3$ was deduced from the HR-ESI-MS and spectroscopic data. Netamine S (5), with two additional H-atoms compared to netamine P (2), was found to be the 2',3'-dihydro analog of 2. The NOESY correlations H–C(3a) (δ (H) 4.25–4.30)/H–C(5a) (2.37–2.45) and H–C(5a)/H–C(7) (1.54–1.60) confirmed the location at the same side of the molecule for these three H-atoms. No correlation was observed between these three H-atoms and H–C(8) (δ (H) 1.92–1.97), excluding placement of H–C(8) at the same side of the molecule.

Biological Studies. The crude extract of the *B. laboutei* sponge showed potent cytotoxic activity against the KB tumor cell line (96.9% inhibition at 10 μ M concentration). Netamines O–Q isolated in sufficient quantities were therefore evaluated for their cytotoxic activities against KB cells. Netamines O and Q were cytotoxic in the range of 10⁻⁵ M. Among the known guanidine compounds bearing a tricyclic 5,6,8b-triazaperhydroacenaphtylene skeleton, cytotoxic activities against cancer cell lines were also observed for ptilocaulin ($\Delta^{8,8a}$), isoptilocaulin ($\Delta^{6,7}$), mirabilin C acetate (pyrimidine), mirabilins F, G, and I ($\Delta^{8,8a}$), mirabilins H and K ($\Delta^{8a,8b}$), and netamines C and D (saturated) [1][4][8][9]. The cytotoxic activity of a synthetic racemic ptilocaulin was studied by *Rubent et al.* [12] in order to evaluate its *in vitro* activities against various cell lines. Moreover, the authors have shown that ptilocaulin was toxic at 50 and 25 mg kg⁻¹ in an *in vivo* L1210 tumor model and was ineffective at lower concentrations (T/C 100–112%).

The crude extract of the *B. laboutei* sponge also displayed promising *in vitro* antiplasmodial activity (IC_{50} 3.26±0.25 µM). Thus, the *in vitro* activity of netamines O–Q was undertaken against *Plasmodium falciparum*. All other compounds were not isolated in sufficient quantities. Netamines O–Q exhibited antimalarial activities with IC_{50} values of 16.99 ± 4.12 , 32.62 ± 3.44 , and 8.37 ± 1.35 µM, respectively (*Table 4*). Four other tricyclic alkaloids have been reported as being active against the malaria parasite *P. falciparum*: a mixture of 1.8a-/8b,3a-dihydro-8 β -hydroxyptilocaulin and 1.8a-/8b,3a-dihydro-8 α -hydroxyptilocaulin (prymidine) with an IC_{50} value of 3.8 µg ml⁻¹ [3], netamine K, and mirabilin A with IC_{50} values of 2.4 and 20.7 µM, respectively.

Table 4. Antiplasmodial Activities of the Crude Extract of B. laboutei and of Netamines O-Q (1-3, resp.) against P. falciparum

Crude extract and pure compounds	<i>IC</i> ₅₀ [µм] ^a)	
Crude extract	3.26 ± 0.25	
1	16.99 ± 4.12	
2	32.62 ± 3.44	
3	8.37 ± 1.35	
Artemisinin ^b)	0.014 ± 0.004	
Chloroquine ^b)	0.050 ± 0.034	

^a) The results were the average means of three replicate determinations \pm SD. ^b) Positive control.

Conclusions. – Five new tricyclic guanidine alkaloids, netamines O-S (1–5; resp.), together with the known compounds netamine E (6) and mirabilin J (7), were isolated from the sponge *B. laboutei*. Netamines O-Q, isolated in sufficient quantities, were evaluated for their cytotoxic and antiplasmodial activities. Netamines O and Q exhibited moderate cytotoxic activities. Netamines O-Q exhibited good antiplasmodial activities.

The authors gratefully acknowledge financial support from the *European Commission* and the *Regional Council of Reunion Island: BIOMOL-TCN program* (Activités Thérapeutiques, Cosmétologiques et Nutraceutiques de MOLécules issues de la BIOdiversité terrestre, marine et microbienne de la zone Sud-Ouest de l'Océan Indien), ERDF (*European Regional Development Fund, Grant No.* 2012– 32698). The authors also express their gratitude to Prof. *M. E. Remanevy* for assistance in sponge collection.

Experimental Part

General. All solvents were anal. or HPLC grade and were used without further purification. Thin layer chromatography (TLC): precoated silica gel $60 F_{254}$ sheets (SiO₂); visualized by UV light at 254 nm and by spraying with vanillin/H₂SO₄ followed by heating. MPLC: Teledyne Isco CombiFlash[®] Companion® with a RediSep prepacked normal-phase column (120 g) and Büchi system, including two C-605 pumps, a C-615 pump manager, a C-660 fraction collector, and a glass column (36 × 46 mm) packed with Macherey-Nagel MN Kieselgel (70-230 µm). Anal. HPLC: Kinetex (4.6 × 100 mm, 5 µm) or Waters SunFire (4.6 × 150 mm, 5 µm) column; Waters 2695 Alliance system equipped with a photodiode array detector (Waters 996), an evaporative light scattering detector (Waters 2420), and a mass spectrometer (Waters Micromass ZQ 2000). Prep. HPLC: Waters SunFire Prep reversed-phase C₁₈ (RP- C_{18}) column (19×150 mm, 5 µm); Waters 600 system controller equipped with a photodiode array detector (Waters 2996). Optical rotations: MCP 300 Modular Circular Polarimeter Anton Paar at 25°. UV Spectra: Varian Cary spectrometer; in MeOH; $\lambda_{max} (\log \varepsilon)$ in nm. IR Spectra: PerkinElmer Spectrum 100 FT-IR spectrometer model instrument; dry film; $\tilde{\nu}$ in cm⁻¹. ¹H- and ¹³C-NMR spectra: Bruker UltraShield Avance-300, -500, and DRX-600 spectrometers; δ in ppm rel. to solvent signals (δ (H) 3.31 and $\delta(C)$ 49.15 for CD₃OD; $\delta(H)$ 8.03 and $\delta(C)$ 163.00 for (D₇)DMF), J in Hz. The spectra were processed using 1D- and 2D-NMR notebook software. NOESY or ROESY spectra were recorded in (D₇)DMF or CD₃OD (see the Supporting Information¹)). CDCl₃ was used only for comparison with known compounds. HR-ESI-MS: LCT Premier XE Micromass spectrometer; in m/z.

Animal Material. The sponge B. laboutei HOOPER, 1996 (phylum, Porifera; class, Demospongiae; order, Poecilosclerida; family, Desmacellidae), identified by J. V., was collected in October 2009 at four sampling stations in Salary Bay, Madagascar: station 1 ($22^{\circ}31'727''$ S, $43^{\circ}13'597''$ E, at 18 m depth), station 2 ($22^{\circ}30'952''$ S, $43^{\circ}12'558''$ E, at 25-30 m depth), station 3 ($22^{\circ}31'988''$ S, $43^{\circ}13'036''$ E, at 30 m depth), station 4 ($22^{\circ}31'822''$ S, $43^{\circ}12'939''$ E, at 25-27 m depth). Seven voucher specimens (# MHNM.16242.1, MHNM.16242.2, MHNM.16242.3, MHNM.16242.4, MHNM.16242.5, MHNM.16242.6, and MHNM.16242.7) were deposited with the Museum d'Histoire Naturelle de Marseille, Palais Longchamp, 1 Bd Philippon, FR-13004 Marseille. Sponge samples were frozen immediately and kept at -20° until processing.

Extraction and Isolation. The CH₂Cl₂/MeOH 1:1 (3×3.5 l, each 24 h) extract (87 g) was obtained by maceration of the freeze-dried sponge (358 g) at r.t. The extract was then subjected to MPLC (SiO₂; isohexane/AcOEt/MeOH with increasing polarity) to give four fractions, *Frs. 1–4. Fr. 1* was eluted with isohexane/AcOEt 95:5, *Fr. 2* was eluted with isohexane/AcOEt 85:15, *Fr. 3* was eluted with AcOEt, and *Fr. 4* was eluted with AcOEt/MeOH 70:30.

¹) Supporting material is available upon request from the authors.

Separation of *Fr.* 4 (2.6 g) by MPLC (*RP-C*₁₈; H₂O/MeOH (+0.1% HCOOH)) gave twelve subfractions, *Frs.* 4.1–4.12. Anal. HPLC (*Waters SunFire* C_{18} (4.6×150 mm, 5 µm); 45% MeOH/H₂O (+ 0.1% HCOOH) to 70% MeOH/H₂O (+0.1% HCOOH), 1 ml min⁻¹ gradient elution over 45 min; UV 254 nm, ELS) analyses showed that the isolated compounds are present in *Frs.* 4.5 (108.8 mg) and 4.10 (150.0 mg). *Fr.* 4.5 was subjected to semi-prep. HPLC (*Waters SunFire RP-C*₁₈ (19×150 mm, 5 µm); 55% MeOH/H₂O (+0.1% HCOOH) to 68% MeOH/H₂O (+0.1% HCOOH), 4.8 ml min⁻¹ gradient elution over 20 min; UV 254 nm) to furnish four subfractions, *Frs.* 4.5.1–4.5.4, containing pure compound **7** (mirabiline J; 5.5 mg). *Fr.* 4.10 was subjected to subsequent semi-prep. HPLC (*Waters SunFire RP-C*₁₈; 75% MeOH/H₂O (+0.1% HCOOH), 5 ml min⁻¹ isocratic elution over 15 min; UV 254 nm) to give five subfractions, *Frs.* 4.10.1–4.10.5. Two of them, *Frs.* 4.10.2 and 4.10.3, afforded pure compounds **6** (netamine E; 4.0 mg) and **3** (netamine Q; 8.9 mg).

Fr. 4 (360 mg) was also subjected to prep. HPLC (*Waters SunFire Prep RP-C₁₈* (19×150 mm, 5 µm); 45% MeOH/H₂O (+0.1% HCOOH) to 70% MeOH/H₂O (+0.1% HCOOH), 17 ml min⁻¹ gradient elution over 45 min; UV 254 nm) to give 13 subfractions, *Frs.* 4.1'-4.13'. One of them afforded **1** (netamine O; 8.4 mg). Three other subfractions were subjected to subsequent prep. HPLC (*Waters SunFire Prep RP-C₁₈*; 55% MeOH/H₂O (+0.1% HCOOH) to 70% MeOH/H₂O (+0.1% HCOOH), 17 ml min⁻¹ gradient elution over 26 min; UV 254 nm) to give pure compounds **2** (netamine P; 3.9 mg), **4** (netamine R; 1.9 mg), and **5** (netamine S; 1.9 mg).

Netamine O (=(3aS*,5aS*,7R*,8R*)-8-[(2Z)-Hex-2-en-1-yl]-3,3a,4,5,5a,6,7,8-octahydro-7-methylcyclopenta[de]quinazolin-2-amine; **1**). Yellow oil. [a]₂₅²⁵ = +61.6 (c=1.33, MeOH). UV: 234 (0.95), 302 (0.28). IR: 2990, 1599. ¹H- and ¹³C-NMR: see *Table 1*. HR-ESI-MS: 274.2271 (M^+ , $C_{17}H_{28}N_3^+$; calc. 274.2278).

Netamine P (=(3aS*,5aS*,7R*,8S*)-8-[(2Z)-Hex-2-en-1-yl]-3,3a,4,5,5a,6,7,8-octahydro-7-propylcyclopenta[de]quinazolin-2-amine; **2**). White solid. [a] $_{D}^{25}$ = +219.0 (c=1.46, MeOH). UV: 235 (0.56). IR: 3290, 1646. ¹H- and ¹³C-NMR: see *Tables 2* and *3*. HR-ESI-MS: 302.2600 (M^+ , $C_{19}H_{32}N_3^+$; calc. 302.2591).

Netamine Q (=(3aS*,5aS*,7S*,8S*)-8-[(2Z)-Hex-2-en-1-yl]-3,3a,4,5,5a,6,7,8-octahydro-7-propylcyclopenta/de]quinazolin-2-amine; **3**). Yellow oil. $[a]_{25}^{25} = +18.3$ (c=0.30, MeOH). UV: 233 (0.94), 305 (0.17). IR: 3300, 1648. ¹H- and ¹³C-NMR: see *Tables 2* and *3*. HR-ESI-MS: 302.2578 (M^+ , $C_{19}H_{32}N_3^+$; calc. 302.2591).

Netamine R (= (3aS*,5aS*,7R*,8S*)-8-Ethyl-3,3a,4,5,5a,6,7,8-octahydro-7-propylcyclopenta[de]quinazolin-2-amine; **4**). Pale yellow oil. [α]_D⁵ = +18.0 (c=0.10, MeOH). UV: 238 (0.63). IR: 3300, 1650. ¹Hand ¹³C-NMR: see *Tables 2* and 3. HR-ESI-MS: 248.2146 (M^+ , $C_{15}H_{26}N_3^+$; calc. 248.2121).

Netamine S (= (3aS*,5aS*,7R*,8S*)-8-*Hexyl*-3,3a,4,5,5a,6,7,8-*octahydro*-7-*propylcyclopenta*[de]*qui-nazolin*-2-*amine*; **5**). Pale brown oil. [a] $_{5}^{5}$ = +20.0 (c = 0.10, MeOH). UV: 248 (0.30). IR: 3300, 1650. ¹H- and ¹³C-NMR: see *Tables 2* and 3. HR-ESI-MS: 304.2748 (M^+ , $C_{19}H_{34}N_3^+$; calc. 304.2747).

Netamine E (= (*3a*\$,*5a*\$,*7***R**,*8*\$)-*8*-*Butyl*-*3*,*3a*,*4*,*5*,*5a*,*6*,*7*,*8*-*octahydro*-*7*-*propylcyclopenta*[de]*quinazolin*-2-*amine*; **6**). Yellow oil. [*a*]_D²⁵ = + 37.0 (*c* = 0.10, CH₂Cl₂). UV: 238 (0.63). HR-ESI-MS: 276.2441 (M^+ , C₁₇H₃₀N₃⁺; calc. 276.2434).

Mirabilin J (=(*3a*\$,*5a*\$,*7*R,8\$)-2-*Amino-8-[(2Z)-hex-2-en-1-yl]-3,3a*,4,5,*5a*,6,*7*8-octahydro-7-methylcyclopenta[de]quinazolin-8-ol; **7**). Brown oil. [α]_D²⁵ = +78.0 (c=0.18, CH₂Cl₂). HR-ESI-MS: 290.2238 (M^+ , C₁₇H₂₈N₃O⁺; calc. 290.2227).

In vitro *Cytotoxicity Assay Against KB Cell Line*. Cell proliferation was measured with *Celltiter 96 Aqueous One* soln. reagent (*Promega*) and results are expressed as the percentage of inhibition of cellular proliferation of KB cells treated for 72 h with 1-3 compared to cells treated with DMSO only (mean \pm SE of triplicate). The *IC*₅₀ determinations were performed in duplicate experiments and are expressed as individual values.

In vitro Antiplasmodial Assays. P. falciparum strains were utilized and details of the assay protocols have been previously reported [13]. Artemisinin (98%; Sigma–Aldrich) and chloroquine (>98%; Sigma–Aldrich) were used as positive controls.

REFERENCES

- [1] G. C. Harbour, A. A. Tymiak, K. L. Rinehart Jr., P. D. Shaw, R. G. Hughes Jr., S. A. Mizsak, J. H. Coats, G. E. Zurenko, L. H. Li, S. L. Kuentzel, J. Am. Chem. Soc. 1981, 103, 5604.
- [2] R. Tavares, D. Daloze, J. C. Braekman, E. Hajdu, R. W. M. Van Soest, J. Nat. Prod. 1995, 58, 1139.
- [3] H.-M. Hua, J. Peng, F. R. Fronczek, M. Kelly, M. T. Hamann, Bioorg. Med. Chem. 2004, 12, 6461.
- [4] W. A. Gallimore, M. Kelly, P. J. Scheuer, J. Nat. Prod. 2005, 68, 1420.
- [5] R. A. Barrow, L. M. Murray, T. K. Lim, R. J. Capon, Aust. J. Chem. 1996, 49, 767.
- [6] A. D. Patil, A. J. Freyer, P. Offen, M. F. Bean, R. K. Johnson, J. Nat. Prod. 1997, 60, 704.
- [7] R. J. Capon, M. Miller, F. Rooney, J. Nat. Prod. 2001, 64, 643.
- [8] M. El-Naggar, M. Conte, R. J. Capon, Org. Biomol. Chem. 2010, 8, 407.
- [9] H. Sorek, A. Rudi, S. Gueta, F. Reyes, M. J. Martin, M. Aknin, E. Gaydou, J. Vacelet, Y. Kashman, *Tetrahedron* 2006, 62, 8838.
- [10] M. Yu, S. S. Pochapsky, B. B. Snider, J. Org. Chem. 2008, 73, 9065.
- [11] E. Gros, A. Al-Mourabit, M.-T. Martin, J. Sorres, J. Vacelet, M. Frederich, M. Aknin, Y. Kashman, A. Gauvin-Bialecki, J. Nat. Prod. 2014, 77, 818.
- [12] R. L. Ruben, B. B. Snider, F. W. Hobbs Jr., P. N. Confalone, B. A. Dusak, *Invest. New Drugs* 1989, 7, 147.
- [13] O. Jansen, M. Tits, L. Angenot, J.-P. Nicolas, P. De Mol, J.-B. Nikiema, M. Frédérich, *Malar. J.* 2012, 11, 289.