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Short Note

Isolation and Characterization of Microsatellite Markers for *Mimusops balata* (Sapotaceae) and Cross-Amplification in Other *Mimusops* Species

Stéphanie Dafreville ^{1,*}, Cláudia Baider ², F. B. Vincent Florens ³, Gérard Lebreton ¹,
Eric Rivière ¹, Dominique Strasberg ⁴ and Marie-Hélène Chevallier ¹

¹ CIRAD, UMR C53 PVBMT, Saint-Pierre 97410, La Réunion, France;

E-Mails: gerard.lebreton@cirad.fr (G.L.); riviere@cirad.fr (E.R.);
marie-helene.chevallier@cirad.fr (M.-H.C.)

² The Mauritius Herbarium, R. E. Vaughan Building, Agricultural Services, Ministry of
Agro-Industry and Food Security, Réduit, Mauritius; E-Mail: clbaider@gmail.com

³ Department of Biosciences, University of Mauritius, Réduit, Mauritius; E-Mail: v.florens@gmail.com

⁴ Université de la Réunion, UMR C53 PVBMT, Saint-Denis 97715, La Réunion, France;
E-Mail: umrc53@univ-reunion.fr

* Author to whom correspondence should be addressed; E-Mail: stephanie.dafreville@cirad.fr;
Tel.: +262-262-499-213 (ext. 123); Fax: +262-262-499-295.

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Abstract: *Mimusops balata* (Sapotaceae) is an endemic tree species from La Réunion and Mauritius. Like many species growing in lowland forests in La Réunion, it has suffered from human disturbances. We developed twelve microsatellite markers for *M. balata* and tested cross-amplification in five other *Mimusops* species to have powerful tools for genetic diversity studies. Genotyping peaks were of very low quality for two loci and were consequently abandoned for the genetic diversity analyses. Ten microsatellite loci were tested on 34 individuals of *M. balata* from two natural populations. The number of alleles per locus ranged from one to seven. The observed and expected heterozygosity levels varied from 0.000 to 0.823, and from 0.000 to 0.812 respectively. Two loci deviated from the Hardy-Weinberg equilibrium. The presence of null alleles was detected for one of these two loci. Nine to ten loci cross-amplified reliably in Mauritian species, for the other three species, four to six loci show successful amplifications. These polymorphic microsatellite

markers are now available for population genetic investigations in *Mimusops* species aiming to establish accurate guidelines for conservation managers.

Keywords: conservation genetics; genetic diversity; La Réunion; Mauritius; nuclear microsatellites

1. Introduction

La Réunion Island belongs to one of the thirty-four global biodiversity hotspots, where high concentration of endemism is undergoing severe loss of habitats [1]. Since the human colonization on the island, 70% of native vegetation have been destroyed or disturbed due to human activities [2]. Lowland forests have been particularly affected by anthropogenic disturbances like habitat destruction, forest fragmentation and logging. Preserving the last remnants of each habitat type on La Réunion is among the main conservation priority. To develop sustainable conservation plans, it is useful to assess information about the structure of genetic diversity and the patterns of gene flow between extant populations to reduce extinction risk [3].

Mimusops balata (Aubl.) C.F. Gaertn. (Sapotaceae) is one of the dominant endemic canopy trees that evolved in lowland forests of Mauritius and La Réunion. Like most of Sapotaceae species occurring on the Mascarene Islands, *M. balata* has an important keystone structure role in these forests. Its natural distribution has been severely contracted and fragmented. Although *M. balata* remained relatively abundant in its habitat [4], investigating its genetic diversity and differentiation are essential to ensure the long-term survival of this species and its functional role in its ecosystem. Here we report the development and characterization of twelve microsatellite markers for *M. balata* from La Réunion Island.

2. Results and Discussion

Twelve potential markers developed through 454 GS-FLX Titanium pyrosequencing were chosen to build a single primer mix for polymorphism analyses in the species *M. balata*. The characteristics of these twelve microsatellite loci are described in the Table 1. Genotyping peaks for the loci MbCIR26 and MbCIR15 were of low quality; allele assignment was therefore very difficult. So polymorphism analyses were not performed for these loci. The other ten markers were used to screen 34 individuals collected from two populations of *M. balata* (Les Makes and Sainte-Thérèse located respectively in the south-west and the south of La Réunion). The number of alleles per locus ranged from one to seven, with an average of 4.1 in Les Makes population and 4.5 in the Sainte-Thérèse population (Table 2). The observed and expected heterozygosities ranged from 0.000 to 0.823, and from 0.000 to 0.812 respectively. There was no evidence for linkage disequilibrium ($p > 0.05$) in any pair of loci. The locus MbCIR65 shows the lowest polymorphism levels. Two loci significantly deviated from Hardy-Weinberg equilibrium (MbCIR4, MbCIR30), suggesting the presence of null alleles, a Wahlund effect or an incomplete sample size. The presence of null alleles was confirmed at the locus MbCIR30.

Table 1. Characteristics of twelve microsatellite loci developed for the tree species *Mimusops balata*.

Locus	Repeat motif	Primer sequence (5' to 3')	Fluorescent Dye	Size range (bp)	Accession number
MbCIR4	(AC) ₁₂	F-TCTGTCATCCCAGTCTGCTG R-TGTTTGATGTGTCTTTAGACCAAG	VIC	89-113	JX413365
MbCIR10	(GT) ₁₃	F-CCGTCTGCACCGTTCTTATT R-TGATTTGCTTAAACTGGGAAGC	6-FAM	132-154	JX413367
MbCIR13	(CA) ₁₄	F-CAGGGATGGTAACAGCTGAC R-TGTTCCAATTACAGGTAAAGGC	NED	137-167	JX413368
MbCIR14	(TG) ₁₄	F-TTTACATTAGATCGGTGCGAG R-AGACCAAAAATAAATACTGGGTATTACA	6-FAM	228-246	JX413375
MbCIR15	(CA) ₁₄	F-ATTATCCCATCAATAATCCCACC R-ATTCTTTTGTCTGTAGTTGTTTACTCA	NED	235-241	JX413376
MbCIR26	(CA) ₁₃	F-AAATACATATAGTGCAACATAACCTGC R-TGTTAGGTAGTGGTAGTCTCTGCTTT	6-FAM	90	JX413374
MbCIR30	(CA) ₁₂	F-GTGTTTTGCTTAAGGGTTGGTC R-CGAACGTGAATTACATGTGTTACC	NED	72-94	JX413373
MbCIR34	(GA) ₁₂	F-AACTAACGGCCAAATAGGCA R-ATACCAGCAGAAAGCCAGCA	PET	92-112	JX413366
MbCIR55	(TC) ₁₂	F-TTGCCTCCTGGAGTAAGTGG R-TCAGACAATTGGCCTTAGCC	PET	158-164	JX413370
MbCIR62	(AG) ₁₁	F-GGACATGTTGCCAATGAGC R-GGGTATCATTAAACTTCTTGGTCA	PET	230-252	JX413371
MbCIR63	(AG) ₁₁	F-CTTGCTGTGTAAGTGGCAACA R-TAGGTTCTCAGTCAGCAACATCT	VIC	234-246	JX413372
MbCIR65	(GA) ₁₁	F-CCAAGGGGCTATTCTCTTC R-CACCCAACGTAAGTCTGCTCA	VIC	149-161	JX413369

Table 2. Genetic diversity revealed by ten microsatellite loci in two natural populations of *Mimusops balata*. N Number of individuals tested; N_A Number of alleles detected; H_O Observed heterozygosity; H_E Expected heterozygosity; Significant deviation from Hardy-Weinberg equilibrium: * $p < 0.05$, *** $p < 0.001$.

Locus	Les Makes (N = 17)			Sainte-Thérèse (N = 17)		
	N _A	H _O	H _E	N _A	H _O	H _E
MbCIR4	7	0.625	0.812 *	7	0.588	0.708
MbCIR10	6	0.765	0.726	6	0.588	0.636
MbCIR13	6	0.588	0.710	6	0.471	0.456
MbCIR14	5	0.687	0.742	5	0.687	0.767
MbCIR30	4	0.294	0.614 *	3	0.000	0.581 ***
MbCIR34	4	0.588	0.533	7	0.823	0.700
MbCIR55	2	0.471	0.426	2	0.529	0.485
MbCIR62	3	0.625	0.523	5	0.823	0.706
MbCIR63	3	0.375	0.329	2	0.118	0.114
MbCIR65	1	0.000	0.000	2	0.062	0.062

The twelve loci were tested for cross-amplification in five other *Mimusops* species (Table 3): *M. petiolaris* (A.DC.) Dubard (MAU; N = 5), *M. erythroxyton* Bojer ex A.DC. (MAU; N = 5), *M. comorensis* Engl. (COM: Comoros; N = 3), *M. coriacea* (A.DC.) Miq. (MAD: Madagascar; N = 3), *M. elengi* L. (IN: India; N = 2). The genotyping peaks for MbCIR26 and MbCIR15 were of low quality too. Nine to ten loci successfully amplified in Mauritian *Mimusops* suggesting that these species are more closely related to *M. balata* than the other species tested in our study for which only four to six loci were successfully transferred. The number of alleles detected with these markers in the congeneric species ranged from one to eight.

Table 3. Cross-species amplification of ten microsatellite markers developed for *Mimusops balata* on species of the genus *Mimusops*. N: Number of individuals tested; N_A: Number of alleles detected; MAU: Mauritius; COM: Comoros; MAD: Madagascar; IN: India.

Locus	N _A (Size range in bp)				
	<i>M. petiolaris</i> MAU (N = 5)	<i>M. erythroxyton</i> MAU (N = 5)	<i>M. comorensis</i> COM (N = 3)	<i>M. coriacea</i> MAD (N = 3)	<i>M. elengi</i> IN (N = 2)
MbCIR4	7 (81–101)	5 (89–109)	4 (87–105)	2 (79–83)	1 (89)
MbCIR10	3 (132–136)	6 (126–150)	3 (136–162)	1 (138)	2 (140–156)
MbCIR13	5 (133–143)	6 (133–153)	3 (125–149)	-	-
MbCIR14	3 (228–240)	-	-	-	-
MbCIR30	5 (76–92)	3 (88–92)	-	-	-
MbCIR34	8 (98–136)	7 (98–136)	-	2 (112–114)	-
MbCIR55	2 (158–162)	4 (144–162)	1 (164)	1 (160)	1 (164)
MbCIR62	3 (246–250)	4 (246–252)	-	1 (234)	-
MbCIR63	4 (224–238)	3 (234–240)	2 (228–232)	2 (238–240)	2 (236–244)
MbCIR65	2 (155–161)	2 (155–161)	-	-	-

3. Experimental Section

DNA was extracted from dried leaf tissue of five individuals of *M. balata* using the DNeasy Plant Mini Kit (Qiagen) and sent to GenoScreen, France [5]. Microsatellite-enriched libraries were developed through 454 GS-FLX Titanium pyrosequencing by the platform GenoScreen (Lille, France) as described in [6]. It was a rapid and low-cost opportunity to develop a library of specific nuclear microsatellite markers for *M. balata*. This technique allowed the identification of 955 potential markers. Twelve dinucleotide microsatellite loci were chosen to build a single primer mix containing 2 µM of each primer (Table 1) for multiplex amplifications. Levels of variation of the twelve microsatellite loci were evaluated among 34 individuals from two populations of *M. balata*: Les Makes (N = 17) and Sainte-Thérèse (N = 17). Multiplex PCR was performed in 15 µL reaction volumes containing 10–20 ng of genomic DNA, 7.5 µL of 2× Type-it Multiplex PCR Master Mix (Qiagen), and 0.2 µM of each primer (in each primer pair the forward primer was fluorescently labeled, see Table 1). PCR reactions were carried out in a GeneAmp PCR System 9700 thermocycler (Applied Biosystems) with the following conditions: 5 min of denaturation at 95 °C, 28 cycles consisting of 30 s of denaturation at 95 °C, 90 s of annealing at 60 °C, and 30 s of extension at 72 °C, and a final extension at 60 °C for 30 min. One microliter of PCR products was combined with 10.7 µL deionized formamide and genotyping was

performed on an ABI PRISM 3130XL Genetic Analyser (Applied Biosystems), using 0.3 μ L of Genescan AB500LIZ size standard (Applied Biosystems). Allele scoring was done using GENEMAPPER version 3.7 software (Applied Biosystems). The number of alleles, observed and expected heterozygosities and deviations from the Hardy-Weinberg equilibrium were calculated and tested with the software GENEPOP version 4.0 [7,8]. The linkage disequilibrium was tested using FSTAT version 2.9.3.2 [9,10] and the presence of null alleles was estimated by MICRO-CHECKER version 2.2.3 [11,12].

4. Conclusions

These results show the potential of these markers for investigating population genetic structure and diversity analyses throughout *M. balata* and closely related *Mimusops* species. Genetic diversity studies can generate helpful information for conservation managers that face the challenge of preserving biodiversity after severe human-induced disturbances.

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References

1. Mittermeier, R.A.; Robles Gil, P.; Hoffman, M.; Pilgrim, J.; Brooks, T.; Mittermeier, C.G.; Lamoureux, J.; da Fonseca, G.A.B. *Hotspots Revisited: Earth's Biologically Richest and Most Endangered Terrestrial Ecoregions*; Conservation International: Washington, DC, USA, 2005; p. 392.
2. Strasberg, D.; Rouget, M.; Richardson, D.M.; Baret, S.; Dupont, J.; Cowling, R.M. An assessment of habitat diversity and transformation on La Réunion Island (Mascarene Islands, Indian Ocean) as a basis for identifying broad-scale conservation priorities. *Biodivers. Conserv.* **2005**, *14*, 3015–3032.
3. Frankham, R. Genetics and extinction. *Biol. Conserv.* **2005**, *126*, 131–140.
4. Florens, F.B.V.; Baider, C.; Martin, G.M.N.; Strasberg, D. Surviving 370 years of human impact: What remains of tree diversity and structure of the lowland wet forests of oceanic island Mauritius? *Biodivers. Conserv.* **2012**, *21*, 2139–2167.
5. GenoScreen Homepage. Available online: <http://www.genoscreen.fr/> (accessed on 6 December 2012).
6. Malausa, T.; Gilles, A.; Megléc, E.; Blanquart, H.; Duthoy, S.; Costedoat, C.; Dubut, V.; Pech, N.; Castagnone-Sereno, O.P.; Délye, C.; *et al.* High-throughput microsatellite isolation through 454 GS-FLX Titanium pyrosequencing of enriched DNA libraries. *Mol. Ecol. Resour.* **2011**, *11*, 638–644.
7. Raymond, M.; Rousset, F. GENEPOP (Version 1.2): Population genetics software for exact tests and ecumenicism. *J. Hered.* **1995**, *86*, 248–249.
8. Genepop on the web. Available online: <http://genepop.curtin.edu.au/> (accessed on 6 December 2012).
9. Goudet, J. FSTAT: A computer program to calculate F-statistics, version 1.2. *J. Hered.* **1995**, *86*, 485–486.

10. FSTAT. Available online: <http://www2.unil.ch/popgen/software/fstat.htm/> (accessed on 6 December 2012).
11. Van Oosterhout, C.; Hutchinson, W.F.; Wills, D.P.M.; Shipley, P. MICRO-CHECKER: Software for identifying and correcting genotyping errors in microsatellite data. *Mol. Ecol. Notes* **2004**, *4*, 535–538.
12. Microsatellite Data Checking Software. Available online: <http://www.microchecker.hull.ac.uk/> (accessed on 6 December 2012).

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